

生物安全型选择标记基因及其应用研究进展

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摘要: 在植物基因工程中, 选择标记基因(selectable marker genes, SMGs)的应用使“真”转化子得以辨认, 但目前常用的抗生素及除草剂类SMGs可能对人类及家畜健康、生态平衡等带来潜在的危害, 这也是影响转基因植物商业化的最重要因素之一。鉴于此, 基于可视化、非生物胁迫、代谢关键酶的安全SMGs迅速崛起, 并成功用于转化子筛选, 且筛选精度与效率明显提高。本文旨在对近年来生物安全型SMGs的有效性及其适用性的研究进展进行分析与综述, 以期对转基因植物的广泛推广奠定基础。

关键词: 转基因植物; 安全标记; 生物安全性

早在20世纪80年代初, 外源基因导入植物基因组的尝试开启了植物育种的新时代, 陆续成功培育了大批抗旱、耐盐碱、抗冻、抗病虫害转基因植物; 在这些转基因植物中, 与外源优异基因相串联的选择标记基因(selectable marker genes, SMGs)多数是抗生素(如*NPTII*、*hpt*)和除草剂类基因(*BAR*、*ARO*、*BXN*、*CSRL*), 其编码的蛋白对抗生素、除草剂或其类似物具有解毒作用, 转化细胞由于含有该类解毒基因而得以生长(Miki和McHugh 2004)。然而, 有些植物如硅藻(*Phaeodactylum tri-cornutum*)对抗生素及除草剂有着天然的抗性(Apt等1996), 且许多海洋藻类生活在高盐环境, 降低了抗生素的活性(Allnut等2000)。此外, 抗生素SMGs可能存在的潜在风险引起人们的广泛关注, 如: (1)当食用转基因食品时, SMGs有可能会发生水平转移致使病原微生物的抗药性增强, 引起抗生素类药物失效, 威胁人类健康; (2)转基因植物进行环境释放时, 可能会由于基因漂移造成物种间的优势关系发生质的改变, 破坏生态环境平衡; (3)在植物生长发育过程中, SMGs的存在可能导致原有基因或外源基因的沉默, 甚至影响二次转化, 如有学者发现用同一个SMG标记几个基因或重复使用同一个启动子驱动不同的SMG, 均有可能导致基因的沉默, 因此存在技术风险(Ebinuma等1997)。鉴于此, 众多研究者将视点转向了基于可视化、非生物胁迫、代谢关键酶的生物安全型SMGs(表1~3), 相较于“第一代标记物”抗生素类及除草剂类基因, 其拥有高安全性、遗传转化高效性及公众认可度高等优势。本文主要对近年来生物安全型SMGs在植物基因工程中的应用进行概述。

1 基于可视化的SMGs

绿色荧光蛋白(green fluorescent protein, GFP)基因(Franklin等2002; Li等2009; Lauersen等2013)、色蛋白基因(Chiang等2014)和生物荧光素酶基因(张秀娟等2013; Lerche等2014)是目前可视化SMGs的代表, 无需施加任何选择压便可筛选出转化细胞(Miki和McHugh 2004)(表1)。GFP是海洋动物维多利亚水母(*Aequorea victoria*)体内的一种光蛋白, 作为视觉标记, 有利于转化细胞特别是活性组织的实时监控(Chalfie等1994)及定位(韩晓敏等2016; 梁会超等2016), 且与“第一代标记物”相比更为精确、高效和安全。Zhu等(2003)将含*GFP-cryIac*融合基因及*GNA*(*glanthis nivalis agglutinin*)基因的双价抗虫表达载体导入烟草(*Nicotiana tabacum*)中, 发现外源基因高效表达, 转基因植株绿色荧光表型与抗虫性密切相关, 简化了抗虫转基因植物筛选程序。*GFP*同抗生素类SMGs筛选效率比较实验显示, *GFP*拥有更为稳定的遗传转化事件(Leclercq等2010), 有力的证据表明用*GFP*鉴定嵌合转基因胡桃(*Juglans regia*)体胚, 其鉴定效率为100%(Escobar等2000)。Gao等(2005)以*GFP*作为SMGs、*TLP*(thaumatin-like protein)为目标基因转化高粱(*Sorghum bicolor*), 通过Southern blot分析获得320株*GFP*表达植株, 后期证明*GFP*的表达与*TLP*存在的相关性为100%。Saika和Toki(2009)发现转化水稻(*Oryza sativa*)时以*GFP*为标记的筛选效率

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表1 基于可视化的安全SMGs在植物遗传转化中的应用

Table 1 The application of visual makers in genetic transformation

标记基因	基因来源	筛选剂	应用物种	参考文献
绿色荧光蛋白基因GFP	维多利亚水母(victoria)	无	高粱(<i>S. bicolor</i>) 胡萝卜(<i>D. carota</i>) 水稻(<i>O. sativa</i>) 橡胶树(<i>Hevea brasiliensis</i>) 拟南芥(<i>A. thaliana</i>) 三角褐指藻(<i>P. tricornutum</i>) 莱茵衣藻(<i>C. reinhardtii</i>)	Gao等2005 Baranski等2006 Saika和Toki 2009 Leclerc等2010 韩晓敏等2016 Zhang等2014 Noor-Mohammadi等2014; Ahmad等2015; Rasala等2012; Oey等2014 Shih等2015
紫色色素蛋白基因shCP	地毯海葵(<i>S. haddoni</i>)	无	眼点拟微绿球藻 (<i>Nannochloropsis oculata</i>)	Lerche等2014
Gaussia分泌型萤光素酶 基因GLUC	海洋桡足类动物 (<i>Gaussia princeps</i>)	无	实球藻(<i>Pandorina morum</i>)	Fuhrmann等2004; Eichler-Stahlberg等2009; Park等2013
海肾萤光素酶基因RLUC	海肾(<i>Renilla reniformis</i>)	无	莱茵衣藻(<i>C. reinhardtii</i>)	

高于hpt。类似情况亦在胡萝卜(*Daucus carota*) (Baranski等2006)、莱茵衣藻(*Chlamydomonas reinhardtii*)、三角褐指藻(*Phaeodactylum tricornutum*)等植物中出现。此外, GFP作为SMG不会导致植物形态变异, 对细胞无毒害作用, 其无需底物诱导表达的优点尤为突出。基于此, GFP是目前广泛用于研究胞间蛋白质运输和体内病毒运动的荧光蛋白(Howard等2004), 但其受宿主叶绿体内源性荧光的影响, 而且需要相应的仪器辅助。Chiang等(2014)发现了一种来源于地毯海葵(*Stichodactyla haddoni*)的GFP类似色蛋白shCP, 该蛋白可被生物降解, 在保障环境与食品安全的同时无需荧光显微技术的辅助, 显色肉眼可见; Shih等(2015)针对其作为遗传转化中的选择标记效率分析表明, 其作为SMGs可有效降低假阳性率。此外, 生物萤光素酶如Gaussia分泌型萤光素酶(gaussia luciferase, GLUC)、海肾萤光素酶(renilla luciferase, RLUC)及萤火虫萤光素酶(firefly luciferase, FLUC)基因作为SMGs的灵敏性要明显高于GFP, 拥有更广的发展潜力(Shah等2003; Zhang等2013)。

2 基于非生物胁迫的SMGs

非生物胁迫如干旱、盐碱、高温等对植物有着致命的破坏。拥有高效抗逆基因的植物可在逆境中生存, 这就预示着其可作为SMGs的可能性(Zhou等2013)。目前, 拟南芥(*Arabidopsis thaliana*)

AtSOS1 (质膜Na⁺/H⁺逆向转运蛋白编码基因)、水稻(*O. sativa*) *OsDREB2A*转录因子、*rstB* (糖基转移酶编码基因)及*MsrB* (蛋氨酸亚砷还原酶B基因)等抗逆基因用于植物遗传转化的可能性相继被证明(表2)。依据高盐诱导*AtSOS1*及干旱、高盐诱导DREB转录因子表达的原理, Zhu等(2008)首次以其为SMGs分别对水稻进行了遗传转化, 发现200 mmol·L⁻¹ NaCl可成功分离得到转化子。Zhang等(2009)将*rstB*作为SMG用于烟草遗传转化, 并以170 mmol·L⁻¹ NaCl为筛选剂, 筛选率高达83.3%。随后*rstB*亦用于紫花苜蓿(*Medicago sativa*)的遗传转化筛选(Zhang等2015)。近期, 有研究发现氧化胁迫基因*MsrB*的超表达在提高植物抗氧化能力的同时亦可用于转化子的筛选, 用甲基紫精(methyl viologen, MV)为筛选剂成功获得了转基因拟南芥和西红柿(*Solanum lycopersicum*) (Li等2012, 2013; Lee等2014, 2015)。本课题组发现共转化霸王(*Zygophyllum xanthoxylum*) *ZxNHX-ZxVPI-1*的百脉根(*Lotus corniculatus*)在200 mmol·L⁻¹ NaCl和缺水状态下生长良好(Bao等2014); 200 mmol·L⁻¹ NaCl和干旱处理下的共转化*ZxNHX-ZxVPI-1*紫花苜蓿生长状况远远优于野生型(Bao等2016; Kang等2016)。由此可见, 抗逆相关基因如*ZxVPI-1*、*ZxNHX*、*ZxH-KTI;1*和*ZxSOS1* (Wu等2011; Ma等2014)均有作为筛选标记的潜能, 可用于植物的遗传转化。

表2 基于非生物胁迫的安全SMGs在植物遗传转化中的应用

Table 2 The application of safety SMGs based on abiotic stress in genetic transformation

标记基因	基因来源	筛选剂	应用物种	参考文献
Na ⁺ /H ⁺ 逆向转运蛋白基因 <i>AtSOS1</i>	拟南芥(<i>A. thaliana</i>)	NaCl	拟南芥(<i>A. thaliana</i>)	Shi等2002
			水稻(<i>O. sativa</i>)	Zhu等2008
DREB转录因子 <i>OsDREB</i>	水稻(<i>O. sativa</i>)	NaCl	水稻(<i>O. sativa</i>)	Zhu等2008
			烟草(<i>N. tabacum</i>)	Zhang等2009
糖基转移酶基因 <i>rstB</i>	大豆根瘤菌RT19 (<i>Sinorhizobium fredii</i> strain RT19)	NaCl	苜蓿(<i>M. sativa</i>)	Zhang等2009, 2015
			拟南芥(<i>A. thaliana</i>)	Li等2012
蛋氨酸亚砷还原酶B基因 <i>MsrB</i>	拟南芥(<i>A. thaliana</i>)	甲基紫精	拟南芥(<i>A. thaliana</i>)	Li等2012
			西红柿(<i>S. lycopersicum</i>)	Li等2013

3 基于代谢关键酶的SMGs

生化代谢是每个生物体必不可少的物质与能量的来源。筛选调控代谢活动的关键酶基因作为转基因工程SMGs已成为许多研究者的首选,因为它们为生物体内固有代谢途径,属绿色选择标记,可有效提高转基因植物为人们所接纳的可能性,具有良好的应用前景。目前,基于糖类代谢、激素代谢、氨基酸代谢途径的SMGs已广泛应用于基因工程(表3)。

3.1 基于糖类代谢的SMGs

植物组织培养中愈伤组织的正常生长分化需要诸如蔗糖、甘露糖、葡萄糖、麦芽糖和糖醇等碳源提供能量。其中甘露糖、木糖、核糖醇和阿拉伯糖醇等不能直接为植物细胞所利用,需要关键酶的代谢转化。采用糖代谢的关键酶基因作为SMGs、相应碳源作为筛选剂具有双馨效果:一是可为转化细胞提供能量获得优势生长,而非转化细胞处于饥饿状态;二是其纯化蛋白没有毒性和致敏性,保证食品安全。

来源于大肠杆菌(*Escherichia coli*)的*manA/pmi*基因作为糖类代谢关键酶基因之一,编码的6-磷酸甘露糖异构酶(6-phosphomannose isomerase, PMI),可将不能直接利用的6-磷酸甘露糖转化为可进入糖酵解途径提供能量的6-磷酸果糖。基于此原理,*manA/pmi*基因标记的离体细胞或组织可转化D-甘露糖从而获得优势增殖,而非转化细胞会因“饥饿”停止生长(Joersbo等1996; Reed等2001)。该体系已成功应用于诸多单子叶和双子叶植物如小麦、水稻、玉米(*Zea mays*)和高粱等重要农作物与经济作物以及豆科牧草中(表3),均获得了较高的筛选效率。*pmi*作为SMGs,在转基因玉米中其低拷贝率

增加了43%~60% (Sivamani等2015);在甘薯(*Ipomoea batatas*)和甘蔗(*Saccharum officinarum*)中应用的效果尤为明显(Mbinda等2015; Zhang等2015)。本课题组以*pmi*为SMGs成功获得了转基因拟南芥和紫花苜蓿,且对紫花苜蓿的遗传转化效率要明显高于*hpt*(未发表数据)。此外,有报告显示,PMI的纯化蛋白没有毒性和致敏性、且转*manA/pmi*基因植株的农艺性状与野生植物相比较没有显著差异(Reed等2001; 卓晓蕾等2007)。当然,*pmi*虽具有以上优点,但仍存在一定短板。有研究显示葡萄(*Vitis vinifera*)胚若长时间在含甘露糖的培养基中生长,其萌发会受到损害(Vaccari和Martinelli 2009);其次,甘露糖并不适用于所有植物如肉桂(*Cinnamomum cassia*)等含内源*pmi*的物种及烟草等对甘露糖不敏感植物(郭倩倩等2011)。

与甘露糖一样,木糖只有在被木糖异构酶(xylose isomerase, XYL)催化为D-木糖时才能被植物细胞所利用。Morawala等(2007)将*xyLA*作为标记成功筛选了油菜(*Brassica napus*)及向日葵(*Helianthus annuus*)的转化体,效率为12%~15%。Chen等(2015)为验证几丁质酶基因启动子的活性将*xyLA*作为SMG、*GUS*为报告基因,成功完成启动子的验证。且有学者发现在对马铃薯的筛选中*xyLA*的筛选效率是*nptII*的10倍(Haldrup等1998)。由上可知,木糖异构酶基因*xyLA*作为SMG潜在能力不可小觑,加之木糖广泛用于食品加工及淀粉工业,其安全性无可非议,相信该类标记基因会越来越实用。此外,也有报道将甘露糖-6-磷酸还原酶(mannose-6-phosphate reductase, M6PR)基因*M6PR*、海藻糖-6-磷酸合成酶1 (trehalose 6 phosphate synthase 1, TPS1)基因*TPS1*、2-DOG-6-磷酸酶(2-DOG-6-P phosphatase,

表3 基于代谢关键酶基因的安全SMGs在植物遗传转化中的应用

Table 3 The application of safety SMGs based on key enzyme of metabolism in genetic transformation

标记分类	标记基因	基因来源	筛选剂	应用物种	参考文献														
基于糖类代谢	6-磷酸甘露糖异构酶基因 <i>pmi</i>	大肠杆菌 (<i>Escherichia coli</i>)	D-甘露糖	大麦(<i>H.vulgare</i>)	Reed等2001														
				玉米(<i>Zea may</i>)	Sivamani等2015														
				硬质小麦(<i>Triticum turgidum</i>)	Gadaleta等2006														
				高粱(<i>S. bicolor</i>)	Gao等2005														
				狗尾草(<i>Pennisetum glaucum</i>)	O’Kennedy等2004														
				百脉根(<i>L. corniculatus</i>)	Guo等2015														
				鹰嘴豆(<i>Cicer arietinum</i>)	Patil等2009														
				豇豆(<i>Vigna unguiculata</i>)	Bakshi等2012														
				李子(<i>Prunus domestica</i>)	Wang等2013a, b														
				黄瓜(<i>Cucumis sativus</i>)	He等2006														
				水稻(<i>O. sativa</i>)	Ho等2003; Paine等2005; Duan等2012; Gui等2014; Hu等2016; Rong等2016														
	木糖异构酶基因 <i>XylA</i>	梭状芽孢杆菌 (<i>Clostridium thermosulfurogenes</i>)	鼠李糖或木糖	玉米(<i>Z. mays</i>)	Guo等2007														
				向日葵(<i>H. annuus</i>)	Morawala等2007														
				油菜(<i>B. napus</i>)	Morawala等2007														
				花生(<i>Arachis hypogaea</i>)	Chen等2015														
				酿酒酵母(<i>Saccharomyces cerevisiae</i>)	Stovicek等2015														
				植物乳杆菌(<i>Lactobacillus plantarum</i>)	Staudigl等2014														
				甘露糖-6-磷酸还原酶 <i>M6PR</i>	芹菜(<i>Apium graveolens</i>)	D-甘露糖	拟南芥(<i>A. thaliana</i>)	Gao和Loescher 2003;											
							烟草(<i>N. tabacum</i>)	Sickler等2007											
								Song等2010											
							海藻糖-6-磷酸合成酶1 <i>AtTPS1</i>	拟南芥(<i>A. thaliana</i>)	高浓度(6%~7%)	拟南芥(<i>A. thaliana</i>)	Avonce等 2004								
2-DOG-6-磷酸酶基因 <i>DOG^α1</i>	酵母(<i>Saccharomyces cerevisiae</i>)	葡萄糖	烟草(<i>N. tabacum</i>)				Leyman等2006												
		脱氧葡萄糖	烟草(<i>N. tabacum</i>)				Kunze等2001												
			马铃薯(<i>S. tuberosum</i>)				Kunze等2001												
阿拉伯糖脱氢酶基因 <i>atlD</i>	大肠杆菌(<i>E. coli</i> strain C)	阿拉伯糖醇	水稻(<i>O. sativa</i>)				LaFayette等2005												
基于激素代谢	异戊烯转移酶基因 <i>ipt</i>	农杆菌(<i>Agrobacterium tumefaciens</i>)	无				烟草(<i>N. tabacum</i>)	Endo等2000											
							水稻(<i>O. sativa</i>)	Endo等2002											
				白杨(<i>Populus tomentosa</i>)	Ebinuma等1997														
				β-葡萄糖醛酸酶基因 <i>uidA/gusA</i>	大肠杆菌(<i>E. coli</i>)	N-3-葡糖苷酸	烟草(<i>N. Tabacum</i>)	Joersbo等1996											
							甜菜(<i>Beta vulgaris</i>)	Snyder等1999											
							发状根基因 <i>Rol</i>	发根农杆菌 (<i>Agrobacterium rhizogenes</i>)	无	烟草(<i>N. tabacum</i>)	Komarmytsky等2004								
										矮牵牛(<i>Petunia hybrida</i>)、	Khan等2010								
										长寿花(<i>Kalanchoe blossfeldiana</i>)	Thirukkumarana等2010; Christensen等2012								
										组氨酸激酶基因 <i>CKII</i>	拟南芥(<i>A. thaliana</i>)	无	拟南芥(<i>A. thaliana</i>)	Zuo等2002					
													促芽转录因子基因 <i>ESR1</i>	拟南芥(<i>A. thaliana</i>)	无	拟南芥(<i>A. thaliana</i>)	Banno等2002		
基于氨基酸代谢	天冬氨酸激酶基因 <i>AK</i>	大肠杆菌(<i>E. coli</i>)	赖氨酸和苏氨酸													马铃薯(<i>S. tuberosum</i>)	Perl等1993		
																大麦(<i>H. vulgave</i>)	Brinch等1999		
																邻氨基苯甲酸合成酶 <i>AS</i>	烟草(<i>N. tabacum</i>)	4-甲基咪唑	烟草(<i>N. tabacum</i>)
				7-甲基DL-色氨酸															2008; Barone等2009;

表3 (续)

标记分类	标记基因	基因来源	筛选剂	应用物种	参考文献
				水稻(<i>O. sativa</i>) 大豆(<i>G. max</i>)	Tsai等2005; Zhang等2015 Yamada等2004 Inaba等2007; Cho等2004
	色氨酸合酶基因 <i>AtTSB1</i>	拟南芥(<i>A. thaliana</i>)	5-甲基-色氨酸	紫云英(<i>Astragalus sinicus</i>) 拟南芥(<i>A. thaliana</i>)	Cho等2004 Daniell等2001; Hsiao等2007
	D-型氨基酸氧化酶基因 <i>DAO1</i>	股博肌红酵母 (<i>Rhodotorula gracilis</i>)	D-丝氨酸和 D-丙氨酸	拟南芥(<i>A. thaliana</i>)、 苹果(<i>M. borkh</i>)	Erikson等2004 Hattasch等2009

2-DOG-6-P)基因 DOG^R1 以及阿拉伯糖脱氢酶(arabitol dehydrogenase)基因 atD 作为SMGs的成功案例(表3)。

3.2 基于激素代谢的SMGs

异戊烯基转移酶(isopentenyl transferase, IPT)是细胞分裂素生物合成第一步的催化酶,也是限速酶,主要催化异戊烯基腺苷-5-单磷酸酯的合成(吴吉林等2005)。基于此,导入 ipt 基因的转化子可在没有外源激素(分裂素、生长素)的培养基上正常增殖分化形成不定芽。在莴苣(*Lactuca sativa*)及烟草转化系统中,其筛选效率明显优于 $nptII$ 基因(Endo等2000)。但 ipt 基因的持续过表达可能会造成植物形态上的畸形(Gan和Amasino 1997; Kang等1999; Mori等2001; Ouwerkerk等2001),通过 ipt 基因融合乙醇诱导型启动子,可有效解决这一缺点并能提高转化效率,该方法已成功的应用于许多物种(Caddick等1998; Roslan等2001)。如Lu等(2010)构建了 $AlcA::ipt$ 和 $35S::ipt$ 载体并转化烟草,结果显示乙醇诱导系统可获得健康叶、花和种子且转化效率提高。另外,这种局限亦可通过SMGs剔除技术(转座子系统、结合位点特异性重组酶系统)克服(Peng等2015; Darwish等2014; Zou等2013)。负性 β -葡萄糖醛酸酶(β -glucuronidase)基因 $gusA/uidA$ 、发状根基因 rol 、组氨酸激酶(histidine kinase)基因 $CKII$ 和促芽转录因子 $ESR1$ 等作为SMGs亦得到了广泛应用(表3)。

3.3 基于氨基酸代谢的SMGs

在氨基酸合成代谢中,末端产物会对关键酶的活性进行反馈抑制或反馈阻遏。以反馈抑制或

阻遏不敏感基因为SMGs,加以相应氨基酸为筛选剂进行转化子筛选的策略,目前已得到了广泛的应用。其中天冬氨酸激酶(aspartate kinase, AK)、邻氨基苯甲酸合成酶(anthranilate synthase, AS)、色氨酸合酶(tryptophansynthase $\beta 1$, TSB1)和D-氨基酸氧化酶(D-amino acidoxidase, DAAO)基因最为科学家们青睐(表3)。

在天冬氨酸代谢途径中,调控关键酶将天冬氨酸盐通过不同支链转换为赖氨酸(lysine, Lys)、苏氨酸(threonine, Thr)、蛋氨酸(methionine, Met)和异亮氨酸(isoleucine, Ile)。有报道指明,毫摩尔级的Lys和Thr便可强烈抑制植物的生长(Arruda等1984; Miao等1988; Rognes等1983),其原因是它们会抑制AK酶的活性从而造成植物Met饥饿。此外,细菌中的AK的敏感性弱于植物中的,所以作为标记的AK主要从植物中提取,具有安全性,已成功应用于烟草(Shaul等1992; Karchi等1993)、马铃薯(*Solanum tuberosum*)和大麦(*Hordeum vulgare*) (Brinch等1999)的遗传转化中。

AS是烟草的内源基因,合成必需氨基酸色氨酸(tryptophan, Trp),它是许多化合物诸如生长素、植保素和次生代谢产物的前体。在莽草酸途径中,分支酸是重要的枢纽物质,Trp就是由分支酸代谢而来。AS是该代谢中的一个关键酶,其将分支酸分解成邻氨基苯甲酸并受Trp的反馈抑制(Singh和Widholm 1974)。AS基因作为SMGs已被用于多种植物(表3)。Barone和Widholm (2008)以烟草为材料,AS为标记,4-甲基吡啶(4MI)或7-甲基DL-色氨酸(7MT)为选择剂,结果发现4MI的选择效率是

7MT的2倍。*AS*基因除了用于核转化, 其亦可用于质体转化(Barone等2009)。但由于该基因与转化植物的原生基因可能有同源性, 造成的转基因沉默会影响转化效率, 这在烟草(Barone和Widholm 2008; Tsai等2005)和大豆(*Glycine max*) (Inaba等2007)中均有报道。因此, *AS*的部署需要进一步改进, 以克服基因沉默。此外, 色氨酸合酶(tryptophan synthase $\beta 1$, *TSB1*)对Trp的合成亦十分重要(Last等1991; Pruitt和Last 1993; Zhao等2001), 可作为SMG。Hsiao等(2007)构建了35S::*AtTSB1*载体用于转化拟南芥, 并以75 $\mu\text{mol}\cdot\text{L}^{-1}$ 5-甲基-色氨酸(5-methyl-tryptophan, 5MT)为筛选剂进行筛选获得了阳性植株。

*DAO 1*基因作为安全SMG的原理在于植物体内少量D-丙氨酸或D-丝氨酸的存在便可毒害细胞, 而*DAO 1*基因编码的DAAO可将其氧化为无毒物质。Erikson等(2004)将从股博肌红酵母(*Rhodotorula gracilis*)中得到的*DAO 1*基因和CaMV 35S启动子整合入拟南芥中, 并且在不同底物如D-丙氨酸或D-丝氨酸下筛选出转基因植株, 同时, 也在转基因烟草、大麦、玉米、云杉上对其进行了测试, 结果与拟南芥一致。同样地, Hattasch等(2009)以*DAO 1*基因为标记获得了转基因苹果。

总之, 生物途径及生物来源SMGs的成功应用, 极大地降低了转基因植株的潜在安全风险, 能够有效安抚公众的消极情绪, 为转基因植物商业化及广泛应用奠定了基础。然而, 随着公众对其安全意识的不断提高, 开发新型生态友好植物来源的SMGs依然是未来生物技术发展的研究重点。

4 展望

生物技术能将异源或同源物种的优良基因快速、准确地整合到植物中, 并通过部署SMGs间接地对性状进行选择。转基因技术中, SMGs的部署十分必要, 利于高效地筛选出目标植株。但抗生素和除草剂类SMGs的使用对人类健康及自然生态系统的潜在不利影响, 迫使科学家们不断挖掘易检测、高效性、多功能的新型生态友好型标记。即便如此, 目前安全SMGs的选择范围依然很狭小, 且存在诸多问题。如*ASA2*的表达可能造成基因的沉默; *pmi*基因的筛选剂D-甘露糖很昂贵; *ipt*基因持续过表达可能会造成植物形态上的畸形

等。开发新的安全SMGs, 尤其是抗逆相关标记迫在眉睫, 它们在作为抗逆基因的同时, 亦可作为标记进行转化子的筛选, 同时这类SMGs来源于植物, 可不必担心引起的基因漂移问题, 通过这种转化一次便可实现多种目的, 是研究人员一直梦寐以求的目标。目前, 具有开发为标记潜能的抗逆基因有DREB转录因子、*rstB*、*MsrB*、*NHX*和*SOS1*。此外, 与“第一代标记物”抗生素类及除草剂类基因相比, 安全SMGs拥有遗传转化高效性, 因为在非转化细胞被抗生素及除草剂杀死的过程中会产生大量的醌类、酚类等毒素, 抑制了转化子的正常生长; 但安全标记SMGs仅是使非转化细胞处于饥饿状态、不会产生其它代谢产物降低转化率。因此, 根据遗传转化及转基因作物商品化发展的趋势, 理想的标记基因研究应当具备安全性、易检测、高效性和多功能性等特点。随着研究的不断深入和完善, 安全SMGs的应用前景十分广阔, 转基因植物的大规模商业化将指日可待。

参考文献

- Ahmad I, Sharma AK, Daniell H, Kumar S (2014). Altered lipid composition and enhanced lipid production in green microalga by introduction of brassica diacylglycerol acyltransferase 2. *Plant Biotechnol J*, 13 (4): 540–550
- Allnut FCT, Kyle DJ, Grossman AR, Apt KE (2000). Methods and tools for transformation of eukaryotic algae. USA, Patent Number 6027900, 2000-02-22
- Apt KE, Kroth-Pancic PG, Grossman AR (1996). Stable nuclear transformation of the diatom *Phaeodactylum tricorutum*. *Mol Gen Genet*, 252 (5): 572–579
- Arruda P, Bright SW, Kueh JSH, Lea PJ, Rognes SE (1984). Regulation of aspartate kinase isoenzymes in barley mutants resistant to lysine plus threonine. *Plant Physiol*, 76 (2): 442–446
- Avonce N, Leyman B, Mascorro-Gallardo JO, Van Dijk P, Thevelein JM, Iturriaga G (2004). The *Arabidopsis* trehalose-6-P synthase *AtTPS1* gene is a regulator of glucose, abscisic acid and stress signaling. *Plant Physiol*, 136 (3): 3649–3659
- Bakshi S, Saha B, Roy NK, Mishra S, Panda SK, Sahoo L (2012). Successful recovery of transgenic cowpea (*Vigna unguiculata*) using the 6-phosphomannose isomerase gene as the selectable marker. *Plant Cell Rep*, 31 (6): 1093–1103
- Banno H, Chua NH (2002). *ESR1-a* plant gene that can promote plant regeneration and transformation. USA, Patent Number 6407312, 2000-06-18
- Bao AK, Wang YW, Xi JJ, Liu C, Zhang JL, Wang SM (2014). Co-expression of xerophyte *Zygophyllum xanthoxylum* *ZxNHX* and *ZxVPI-1* enhances salt and drought tolerance in transgenic *Lotus corniculatus* by increasing cations accumulation. *Funct Plant*

- Biol, 41 (2): 203–214
- Bao AK, Du BQ, Touil L, Kang P, Wang QL, Wang SM (2016). Co-expression of tonoplast Cation/H⁺ antiporter and H⁺-pyrophosphatase from xerophyte *Zygophyllum xanthoxylum* improves alfalfa plant growth under salinity, drought and field conditions. *Plant Biotechnol J*, 14: 964–975
- Baranski R, Klocke E, Schumann G (2006). Green fluorescent protein as an efficient selection marker for *Agrobacterium rhizogenes* mediated carrot transformation. *Plant Cell Rep*, 25 (3): 190–197
- Barone P, Widholm JM (2008). Use of 4-methylindole or 7-methyl-DL-tryptophan in a transformant selection system based on the feedback insensitive anthranilate synthase α -subunit of tobacco (*ASA2*). *Plant Cell Rep*, 27 (3): 509–517
- Barone P, Zhang XH, Widholm JM (2009). Tobacco plastid transformation using the feedback insensitive anthranilate synthase [α]-subunit of tobacco (*ASA2*) as a new selectable marker. *J Exp Bot*, 60 (11): 3195–3202
- Brinch-Pedersen H, Olsen O, Knudsen S, Holm PB (1999). An evaluation of feed-back insensitive aspartate kinase as a selectable marker for barley (*Hordeum vulgare* L.) transformation. *Hereditas*, 131 (3): 239–245
- Caddick MX, Greenland AJ, Jepson I, Krause KP, Qu N, Riddell KV, Salter MG, Schuch W, Sonnewald U, Tomsett AB (1998). An ethanol-inducible gene switch for plants: manipulation of carbon metabolism. *Nat Biotechnol*, 16 (2): 177–180
- Chalfie M, Tu Y, Euskirchen G, Ward WW, Prasher DC (1994). Green fluorescent protein as a marker for gene expression. *Science*, 263 (5148): 151
- Chen XL, Song RT, Yu MY, Sui JM, Wang JS, Qiao LX (2015). Cloning and functional analysis of the chitinase gene promoter in peanut. *Genet Mol Res Gmr*, 14 (4): 12710–12722
- Chiang CY, Chen YL, Tsai HJ (2014). Different visible colors and green fluorescence were obtained from the mutated purple chromoprotein isolated from sea anemone. *Mar Biotechnol*, 16 (4): 436–446
- Cho HJ, Brotherton JE, Widholm JM (2004). Use of the tobacco feed-back-insensitive anthranilate synthase gene (*ASA2*) as a selectable marker for legume hairy root transformation. *Plant Cell Rep*, 23 (1-2): 104–113
- Christensen B, Sriskandarajah S, Muller R, Serek M (2012). Quality improvement of ornamental plants by introducing dwarfism into *Kalanchoe blossfeldiana*. *Acta Hort*, 941 (941): 109–116
- Daniell H, Wiebe PO, Fernandez-San Milan A (2001). Antibiotic-free chloroplast genetic engineering - an environmentally friendly approach. *Trends Plant Sci*, 6 (6): 237–239
- Duan Y, Zhai C, Li H, Li J, Mei W, Gui H, Ni D, Song F, Li L, Zhang W, Yang J (2012). An efficient and high-throughput protocol for *Agrobacterium*-mediated transformation based on phosphomannose isomerase positive selection in *Japonica* rice (*Oryza sativa* L.). *Plant Cell Rep*, 31 (9): 1611–1624
- Ebinuma H, Sugita K, Matsunaga E, Yamakido M (1997). Selection of marker-free transgenic plants using the isopentenyl transferase gene. *Proc Natl Acad Sci USA*, 94 (6): 2117–2121
- Eichler-Stahlberg A, Weisheit W, Ruecker O, Heitzer M (2009). Strategies to facilitate transgene expression in *Chlamydomonas reinhardtii*. *Planta*, 229 (4): 873–883
- Endo S, Kasahara T, Sugita K (2000). The isopentenyl transferase gene is effective as a selectable marker gene for plant transformation into tobacco (*Nicotiana tabacum* cv. Petite Havana SRI). *Plant Cell Rep*, 20 (1): 60–66
- Endo S, Sugita K, Sakai M, Tanaka H, Ebinuma H (2002). Single-step transformation for generating marker-free transgenic rice using the *ipt*-type MAT vector system. *Plant J*, 30 (1): 115–122
- Erikson O, Hertzberg M, Nasholm T (2004). A conditional marker gene allowing both positive and negative selection in plants. *Nat Biotechnol*, 22 (4): 455–458
- Escobar MA, Park JI, Polito VS, Leslie CA, Uratsu SL, McGranahan GH, Dandekar AM (2000). Using GFP as a scorable marker in Walnut somatic embryo transformation. *Ann Bot-London*, 85 (6): 831–835
- Franklin S, Ngo B, Efué E, Mayfield SP (2002). Development of a GFP reporter gene for *Chlamydomonas reinhardtii* chloroplast. *Plant J*, 30 (6): 733–744
- Fuhrmann M, Hausherr A, Ferbitz L, Schodl T, Heitzer M, Hegemann P (2004). Monitoring dynamic expression of nuclear genes in *Chlamydomonas reinhardtii* by using a synthetic luciferase reporter gene. *Plant Mol Biol*, 55 (6): 869–881
- Gadaleta A, Giancaspro A, Blechl A, Blanco A (2006). Phosphomannose isomerase, *pmi*, as a selectable marker gene for durum wheat transformation. *J Cereal Sci*, 43 (1): 31–37
- Gan S, Amasino RM (1997). Making sense of senescence: molecular genetic regulation and manipulation of leaf senescence. *Plant Physiol*, 113 (2): 313–319
- Gao XQ, Zhou J, Li J, Zou XW, Zhao JH, Li QL, Xia R, Yang RF, Wang DK, Zuo ZX, et al (2015). Efficient generation of marker-free transgenic rice plants using an improved transposon-mediated transgene reintegration strategy. *Plant Physiol*, 167 (1): 11–24
- Gao Z, Jayaraj J, Muthukrishnan S, Claffin L, Liang GH (2005). Efficient genetic transformation of sorghum using a visual screening marker. *Genome*, 48 (2): 321–333
- Gao Z, Loescher WH (2003). Expression of a celery mannose-6-phosphate reductase in *Arabidopsis thaliana* enhances salt tolerance and induces biosynthesis of both mannitol and a glucosyl-mannitol dimer. *Plant Cell Environ*, 26 (2): 275–283
- Gao Z, Xie X, Ling Y, Muthukrishnan S, Liang GH (2005). *Agrobacterium tumefaciens*-mediated sorghum transformation using a mannose selection system. *Plant Biotechnol J*, 3 (6): 591–599
- Gui HP, Li X, Liu YB, Han K, Li XG (2014). The relationship between *PMI* (*manA*) gene expression and optimal selection pressure in indica rice transformation. *Plant Cell Rep*, 33 (7): 1081–1090
- Guo QQ, Ma JT, Yuan B, Zhou ML, Wu YM (2015). High-efficiency *Agrobacterium*-mediated transformation of *Lotus corniculatus* L. using phosphomannose isomerase positive selection. *Plant Cell Tiss Org*, 121 (2): 1–10
- Guo QQ, Sun XS, Tang YX, Wu YM (2011). Applications of phosphomannose isomerase based selection system in transgenic

- plants. *J Agr Sci Tech*, 13 (6): 12–19 (in Chinese with English abstract) [郭倩倩, 孙熙森, 唐益雄, 吴燕民(2011). 新型筛选标记磷酸甘露糖异构酶基因在转基因植物中的应用. *中国农业科技导报*, 13 (6): 12–19]
- Guo XM, Zhang XD, Liang RQ, Zhang LQ, Chen YF (2007). Maize transformation using xylose isomerase gene as a selection marker. *J Plant Physiol Mol Biol*, 33 (6): 547–552 (in Chinese with English abstract) [郭新梅, 张晓东, 梁荣奇, 张立全, 陈耀锋(2007). 以木糖异构酶基因为筛选标记的玉米遗传转化. *植物生理与分子生物学学报*, 33 (6): 547–552]
- Haldrup A, Petersen SG, Okkels FT (1998). Positive selection: a plant selection principle based on xylose isomerase, an enzyme used in the food industry. *Plant Cell Rep*, 18 (1): 76–81
- Han XM, Xing DD, Yang Q, Li GJ, Wang RG (2016). Cloning and expression analysis of *CiMYB177* from *Caragana intermedia*. *Plant Physiol J*, 52 (5): 635–644 (in Chinese with English abstract) [韩晓敏, 邢丹丹, 杨杞, 李国婧, 王瑞刚(2016). 中间锦鸡儿*CiMYB177*的克隆及表达分析. *植物生理学报*, 52 (5): 635–644]
- Hattasch CH, Flachowsky H, Hanke MV (2009). Evaluation of an alternative D-amino acid/DAAO selection system for transformation in apple (*Malus domestica* Borkh.). *J Hort Sci Biotech*, 1 (1): 188–194
- He Z, Duan ZZ, Liang W, Chen F, Yao W, Liang HW, Yue CY, Sun ZX, Chen F, Dai JW (2006). Mannose selection system used for cucumber transformation. *Plant Cell Rep*, 25 (9): 953–958
- Hoa TT, Al-Babili S, Schaub P, Potrykus I, Beyer P (2003). Golden Indica and Japonica rice lines amenable to deregulation. *Plant Physiol*, 133 (1): 161–169
- Howard AR, Heppler ML, Ju HJ, Krishnamurthy K, Payton ME, Verchot-Lubicz J (2004). *Potato virus X* TGBp1 induces plasmodesmata gating and moves between cells in several host species whereas CP moves only in *N. benthamiana* leaves. *Virology*, 328 (2): 185–197
- Hsiao P, Sanjaya, Su RC, da Silva JAT, Chan MT (2007). Plant native tryptophan synthase beta 1 gene is a non-antibiotic selection marker for plant transformation. *Planta*, 225 (4): 897–906
- Hu L, Li H, Qin RY, Xu RF, Li J, Li L, Wei PC, Yang JH (2016). Plant phosphomannose isomerase as a selectable marker for rice transformation. *Sci Rep-UK*, 6 (1): 25921–25930
- Inaba Y, Brotherton JE, Ulanov A, Widholm JM (2007). Expression of a feedback insensitive anthranilate synthase gene from tobacco increases free tryptophan in soybean plants. *Plant Cell Rep*, 26 (10): 1763–1771
- Joersbo M, Okkels FT (1996). A novel principle for selection of transgenic plant cells: positive selection. *Plant Cell Rep*, 16 (3/4): 219–221
- Kang P, Bao AK, Kumar T, Pan YQ, Bao ZLT, Wang F, Wang SM (2016). Assessment of stress tolerance, productivity, and forage quality in T1 transgenic alfalfa co-overexpressing *ZxNHX* and *ZxVPI-1* from *Zygothlyllum xanthoxylum*. *Front Plant Sci*, 7 (1): 1598–1608
- Kang HG, Fang YW, Singh KB (1999). A glucocorticoid-inducible-transcription system causes severe growth defects in *Arabidopsis* and induces defense-related genes. *Plant J*, 20 (1): 127–133
- Karchi H, Shaul O, Galili G (1993). Seed-specific expression of a bacterial desensitized aspartate kinase increases the production of seed threonine and methionine in transgenic tobacco. *Plant J*, 3 (5): 721–727
- Keravala A, Groth AC, Jarrahan S, Thyagarajan B, Hoyt JJ, Kirby PJ, Calos MP (2006). A diversity of serine phage integrases mediate site-specific recombination in mammalian cells. *Mol Genet Genomics*, 276 (2): 135–146
- Khan RS, Thirukkumaran G, Nakamura I, Mii M (2010). *Rol* (root loci) genes as positive selection marker to produce marker-free *Petunia hybrida*. *Plant Cell Tiss Org Cult*, 101 (3): 279–285
- Komarnitsky S, Gaume A, Garvey A, Borisjuk N, Raskin I (2004). A quick and efficient system for antibiotic-free expression of heterologous genes in tobacco roots. *Plant Cell Rep*, 22 (10): 765–773
- Kunze I, Ebner M, Heim U, Geiger M, Sonnewald U, Herbers K (2001). 2-Deoxyglucose resistance: a novel selection marker for plant transformation. *Mol Breeding*, 7 (3): 221–227
- LaFayette PR, Kane PM, Phan BH, Parrott WA (2005). Arabitol dehydrogenase as a selectable marker for rice. *Plant Cell Rep*, 24 (1): 596–602
- Last RL, Bissinger PH, Mahoney DJ, Radwanski ER, Fink GR (1991). Tryptophan mutants in *Arabidopsis*: the consequences of duplicated tryptophan synthase beta genes. *Plant Cell*, 3 (1): 345–358
- Lauersen KJ, Berger H, Mussgnug JH, Kruse O (2013). Efficient recombinant protein production and secretion from nuclear transgenes in *Chlamydomonas reinhardtii*. *J Biotechnol*, 167 (2): 101–110
- Leclercq J, Lardet L, Martin F, Chapuset T, Oliver G, Montoro P (2010). The green fluorescent protein as an efficient selection marker for *Agrobacterium tumefaciens*-mediated transformation in *Hevea brasiliensis* (Müll. Arg). *Plant Cell Rep*, 29 (1): 513–522
- Lee SH, Li CW, Koh KW, Chuang HY, Chen YR, Lin CS, Chan MT (2014). *MSRB7* reverses oxidation of GSTF2/3 to confer tolerance of *Arabidopsis thaliana* to oxidative stress. *J Exp Bot*, 65 (17): 5049–5062
- Lee SH, Li CW, Liao CH, Chang PY, Liao LJ, Lin CS, Chan MT (2015). Establishment of an *Agrobacterium*-mediated genetic transformation procedure for the experimental model orchid *Erycina pusilla*. *Plant Cell Tiss Org Cult*, 120 (1): 211–220
- Lerche K, Hallmann A (2014). Stable nuclear transformation of *Pandorina morum*. *Bmc Biotechnol*, 14 (1): 1–16
- Leyman B, Avonce N, Ramon M, Dijk PV, Iturriaga G, Thevelein JM (2006). Trehalose-6-phosphate synthase as an intrinsic selection marker for plant transformation. *J Biotechnol*, 121 (3): 309–317
- Li CW, Lee SH, Chan MT (2013). Utilization of the plant methionine sulfoxide reductase B genes as selectable markers in *Arabidopsis* and tomato transformation. *Plant Cell Tiss Org Cult*, 113 (3): 555–563
- Li CW, Lee SH, Chieh PS, Lin CS, Wang YC, Chan MT (2012). *Arabidopsis* root abundant cytosolic methionine sulfoxide reductase B genes *MsrB7* and *MsrB8* are involved in tolerance to oxidative stress. *Plant Cell Physiol*, 53 (10): 1707–1719

- Li SS, Tsai HJ (2009). Transgenic microalgae as a non-antibiotic bactericide producer to defend against bacterial pathogen infection in the fish digestive tract. *Fish Shellfish Immun*, 26 (2): 316–325
- Liang HC, Gao LL, Hu ZF, Gong Ti, Chen JJ, YANG JL, Zhu P (2016). Expression, subcellular localization and characterization of dammarediol-II synthase of *Panax ginseng* in *Saccharomyces cerevisiae*. *Acta Pharm Sin*, 2016 (6): 998–1003 (in Chinese with English abstract) [梁会超, 高丽丽, 胡宗凤, 巩婷, 陈晶晶, 杨金玲, 朱平(2016). 人参达玛烯二醇-II合酶在酿酒酵母中的表达、定位及功能研究. *药理学报*, 2016 (6): 998–1003]
- Lu L, Zhu YY, Liu Y, Zhao DG (2010). Ethanol inducible isopentenyl transferase as a high efficiency marker for tobacco transformation. *Afr J Biotechnol*, 9 (48): 8139–8145
- Ma Q, Li YX, Yuan HJ, Hu J, Wei L, Bao AK, Zhang JL, Wang SM (2014). ZxSOS1 is essential for long-distance transport and spatial distribution of Na⁺ and K⁺ in the xerophyte *Zygophyllum xanthoxylum*. *Plant Soil*, 374 (1-2): 661–676
- Mbinda W, Nawiri S, Nzaro M, Kinyagia B, Mgtutu A, Oduor R (2015). Evaluation of the phosphomannose isomerase-based selection system for genetic transformation of sweetpotato. *Inter J Life Sci*, 9 (3): 46–53
- Miao S, Duncan DR, Widholm JM (1988). Selection of regenerable maize callus cultures resistant to 5-methyl-DL-tryptophane, S-2-aminoethyl-L-cysteine and high levels of L-lysine plus L-threonine. *Plant Cell Tiss Org Cult*, 14 (14): 3–14
- Miki B, McHugh S (2004). Selectable marker genes in transgenic plants: applications, alternatives and biosafety. *J Biotechnol*, 107 (3): 193–232
- Morawala PV, Rajyashri KR (2007). Novel efficient transformation method for sunflower and oil seeds based on positive selection. USA, Patent Number US 2009/0288230 A1, 2009-11-19
- Mori M, Fujihara N, Mise K, Furusawa I (2001). Inducible high level mRNA amplification system by viral replicase in transgenic plants. *Plant J*, 27 (1): 79–86
- Noor-Mohammadi S, Pourmir A, Johannes TW (2014). Method for assembling and expressing multiple genes in the nucleus of microalgae. *Biotechnol Lett*, 36 (3): 561–566
- O'Kennedy MM, Burger JT, Botha FC (2004). Pearl millet transformation system using the positive selectable marker gene phosphomannose isomerase. *Plant Cell Rep*, 22 (9): 684–690
- Oey M, Ross IL, Hankamer B (2014). Gate way-assisted vector construction to facilitate expression of foreign proteins in the chloroplast of single celled algae. *PLoS One*, 9 (2): e86841–e86844
- Ouwerkerk PBF, de Kam RJ, Hoge JHC, Meijer AH (2001). Glucocorticoid-inducible gene expression in rice. *Planta*, 213 (3): 370–378
- Paine JA, Shipton CA, Chaggar S, Howells RM, Kennedy MJ, Vernon G, Wright SY, Hinchliffe E, Adams JL, Silverstone AL, Drake R (2005). Improving the nutritional value of golden rice through increased pro-vitamin A content. *Nat Biotechnol*, 23 (4): 482–487
- Park S, Lee Y, Lee JH, Jin ES (2013). Expression of the high light-inducible dunaliella LIP promoter in *Chlamydomonas reinhardtii*. *Planta*, 238 (6): 1147–1156
- Patil G, Deokar A, Jain PK, Thengane RJSR (2009). Development of a phosphomannose isomerase-based *Agrobacterium*-mediated transformation system for chickpea (*Cicer arietinum* L.). *Plant Cell Rep*, 28 (11): 1669–1676
- Perl A, Galili S, Shaul O, Ben-Tzvi I, Galili G (1993). Bacterial dihydrodipicolinate synthase and desensitized aspartate kinase: two novel selectable markers for plant transformation. *Nat Biotechnol*, 11 (6): 715–718
- Pruitt KD, Last RL (1993). Expression patterns of duplicate tryptophan synthase beta genes in *Arabidopsis thaliana*. *Plant Physiol*, 102 (1): 1019–1026
- Rasala BA, Lee PA, Shen Z, Briggs SP, Mendez M, Mayfield SP (2012). Robust expression and secretion of Xylanase1 in *Chlamydomonas reinhardtii* by fusion to a selection gene and processing with the FMDV 2A peptide. *PLoS One*, 7 (8): e43349–e43359
- Reed J, Privalle L, Powell ML, Meghji M, Dawson J, Dunder E, Suttie J, Wenck A, Launis K, Kramer C, et al (2001). Phosphomannose isomerase: an efficient selectable marker for plant transformation. *In Vitro Cell Dev-Pl*, 37 (37): 127–132
- Rognes SE, Simon WJ, Mifflin BJ (1983). Feedback-insensitive aspartate kinase isoenzymes in barley mutants resistant to lysine plus threonine. *Planta*, 157 (1): 32–38
- Rong RJ, Wu PC, Lan JP, Wei HF, Wei J, Chen H, Shi JN, Hao YJ, Liu LJ, Dou J, et al (2016). Western blot detection of PMI protein in transgenic rice. *J Integr Agr*, 15 (4): 726–734
- Roslan HA, Salter MG, Wood CD, White MRH, Croft KP, Robson F, Coupland G, Doonan J, Laufs P, Tomsett AB, Caddick MX (2001). Characterization of the ethanol-inducible alc gene-expression system in *Arabidopsis thaliana*. *Plant J*, 28 (2): 225–235
- Saika H, Toki S (2009). Visual selection allows immediate identification of transgenic rice calli efficiently accumulating transgene products. *Plant Cell Rep*, 28 (4): 619–626
- Shah K, Tang Y, Breakefield X, Weissleder R (2003). Real-time imaging of TRAIL-induced apoptosis of glioma tumors in vivo. *Oncogene*, 22 (44): 6865–6872
- Shaul O, Galili G (1992). Threonine overproduction in transgenic tobacco plants expressing a mutant desensitized aspartate kinase of *Escherichia coli*. *Plant Physiol*, 100 (3): 1157–1163
- Shi HZ, Lee BH, Wu SJ, Zhu JK (2002). Overexpression of a plasma membrane Na⁺/H⁺ antiporter gene improves salt tolerance in *Arabidopsis thaliana*. *Nat Biotechnol*, 21 (1): 81–85
- Shih CH, Chen HY, Lee HC, Tsai HJ (2015). Purple chromoprotein gene serves as a new selection marker for transgenesis of the microalga *Nannochloropsis oculata*. *PLoS One*, 10 (3): e0120780–e0120796
- Sickler CM, Edwards GE, Kiirats O, Gao Z, Loescher W (2007). Response of mannitol-producing *Arabidopsis thaliana* to abiotic stress. *Funct Plant Biol*, 34 (4): 382–391
- Singh M, Widholm JM (1974). Measurement of the five enzymes which convert chorismate to tryptophan in wheat plants (*Triticum aestivum* L.). *Physiol Plant*, 32 (3): 240–246
- Snyder GW, Ingersoll JC, Smigocki AC, Owens LD (1999). Introduction of pathogen defense genes and a cytokinin biosynthesis gene into sugarbeet (*Beta vulgaris* L.) by *Agrobacterium* or particle bombardment. *Plant Cell Rep*, 18 (10): 829–834

- Song G, Sink KC, Ma Y, Herlache T, Hancock JF, Loescher WH (2010). A novel mannose-based selection system for plant transformation using celerymannose-6-phosphatereductase gene. *Plant Cell Rep*, 29 (2): 163–172
- Staudigl P, Haltrich D, Peterbauer CK (2014). L-Arabinose isomerase and D-Xylose isomerase from *Lactobacillus reuteri*: characterization, co-expression in the food grade host *Lactobacillus plantarum*, and application in the conversion of D-galactose and D-glucose. *J Agr Food Chem*, 62 (7): 1617–1624
- Stovicek V, Borja GM, Forster J, Borodina I (2015). EasyClone 2.0: expanded toolkit of integrative vectors for stable gene expression in industrial *Saccharomyces cerevisiae* strains. *J Ind Microbiol Biot*, 42 (11): 1519–1531
- Thirukkumara G, Ntui VO, Khan RS, Nakamura I, Mii M (2010). Generation of phenotypically normal marker-free transgenic plants of *Kalanchoe blossfeldiana* through hairy root induction. *Plant Biotechnol-GA*, 27 (2): 147–153
- Tsai FY, Brotherton JE, Widholm JM (2005). Overexpression of the feedback-insensitive anthranilate synthase gene in tobacco causes tryptophan accumulation. *Plant Cell Rep*, 23 (8): 548–556
- Vaccari I, Martinelli L (2009). Evaluation of the phosphomannose isomerase-based selection system for gene transfer in grape. *Vitis*, 48 (3): 137–144
- Wang H, Petri C, Burgos L, Albuquerque N (2013a). Phosphomannose-isomerase as a selectable marker for transgenic plum (*Prunus domestica* L.). *Plant Cell Tiss Org Cult*, 113 (2): 189–197
- Wang Y, Chen B, Hu Y, Li J, Lin Z (2015). Inducible excision of selectable marker gene from transgenic plants by the cre/lox site-specific recombination system. *Transgenic Res*, 14 (5): 605–614
- Wang Y, Hua W, Wang J, Hannoufa A, Xu Z, Wang Z (2013b). Deep sequencing of *Lotus corniculatus* L. reveals key enzymes and potential transcription factors related to the flavonoid biosynthesis pathway. *Mgg Mol Gen Genet*, 288 (3-4): 131–139
- Wu GQ, Xi JJ, Wang Q, Bao AK, Ma Q, Zhang JL, Wang SM (2011). The *ZxNHX* gene encoding tonoplast Na^+/H^+ antiporter from the xerophyte *Zygophyllum xanthoxylum* plays important roles in response to salt and drought. *J Plant Physiol*, 168 (1): 758–767
- Wu JL, Wang ZH, Ye QS, Li L (2005). A review of the advances in cytokinin biosynthesis *ipt* Gene. *Subtrop Plant Sci*, 34 (2): 66–69 (in Chinese with English abstract) [吴吉林, 王再花, 叶庆生, 李玲(2005). 细胞分裂素合成基因*ipt*研究进展. 亚热带植物科学, 34 (2): 66–69]
- Yamada T, Tozawa Y, Hasegawa H, Terakawa T, Ohkawa Y, Wakasa K (2005). Use of the feedback-insensitive α -subunit of anthranilate synthase as a selectable marker for transformation of rice and potato. *Mol Breeding*, 14 (14): 363–373
- Yu Y, Song WK, Liu CY, Gao YL, Li WF, Sun DJ, Chen QS, Hu GH (2008). Research development of key enzymes gene on aspartic acid metabolic pathy way in plant. *Biotechnol Bull*, 2008 (s1): 7–11 (in Chinese with English abstract) [于妍, 宋万坤, 刘春燕, 高运来, 李文福, 孙殿君, 陈庆山, 胡国华(2008). 植物天冬氨酸代谢途径关键酶基因研究进展. 生物技术通报, 2008 (s1): 7–11]
- Zhang C, HuH (2014). High-efficiency nuclear transformation of the diatom *Phaeodactylum tricornutum* by electroporation. *Mar Genom*, 6 (1): 63–66
- Zhang MQ, ZhuoXL, Wang JH, Wu Y, Yao W, Chen RK (2015). Effective selection and regeneration of transgenic sugarcane plants using positive selection system. *In Vitro Cell Dev-PI*, 51 (1): 52–61
- Zhang WJ, Wang T (2015). Enhanced salt tolerance of alfalfa (*Medicago sativa*) by *rstB* gene transformation. *Plant Sci*, 234 (1): 110–118
- Zhang WJ, Yang SS, Shen XY, Jin YS, Zhao HJ, Wang T (2009). The salt-tolerance gene *rstB* can be used as a selectable marker in plant genetic transformation. *Mol Breeding*, 23 (2): 269–277
- Zhang XH, Brotherton JE, Widholm JM (2015). Co-expression of the tobacco anthranilate synthase β -subunit with its feedback-insensitive α -subunit as a selectable marker that also markedly increases the free tryptophan content. *In Vitro Cell Dev-PI*, 51 (5): 564–570
- Zhang XJ, Huang H, Xie PW, Wang XJ, Wang SQ (2013). Research progress and application of the secreted gaussia luciferase. *Lett Biotechnol*, 24 (5): 722–726 (in Chinese with English abstract) [张秀娟, 黄海, 谢佩雯, 王学军, 王升启(2013). Gaussia分泌型萤光素酶的研究进展及其应用. 生物技术通讯, 24 (5): 722–726]
- Zhao Y, Christensen SK, Fankhauser C, Cashman JR, Cohen JD, Weigel D, Chory J (2001). A role for flavin monooxygenase like enzymes in auxin biosynthesis. *Science*, 291 (1): 306–309
- Zhou M, Li DY, Li ZG, Hu Q, Yang CH, Zhu LH, Luo H (2013). Constitutive expression of a miR319 gene alters plant development and enhances salt and drought tolerance in transgenic creeping bent grass. *Plant Physiol*, 161 (3): 1375–1391
- Zhu SW, Qin HM, Sun JS, Tian YC (2003). Application of GFP gene in the study of insect resistant transgenic plants. *Acta Bot Sin*, 45 (6): 654–658
- Zhu ZG, Wu R (2008). Regeneration of transgenic rice plants using high salt for selection without the need for antibiotics or herbicides. *Plant Sci*, 174 (5): 519–523
- Zuo JR, Niu QW, Ikeda Y, Chua NH (2002). Marker-free transformation: increasing transformation frequency by the use of regeneration promoting genes. *Curr Opin Biotech*, 13 (2): 173–180
- Zuo XL (2007). Study on the transformation and risk assessment of *PMI* gene in sugarcane [Master's thesis]. Fujian: Fujian Agriculture and Forestry University (in Chinese with English abstract) [卓晓蕾(2007). 甘蔗遗传转化中的*pmi*筛选标记及其安全性评价(硕士学位论文). 福州: 福建农林大学]

Research progress of biosafety selectable marker genes

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Abstract: In genetic engineering, selectable marker genes (SMGs) are generally deployed to identify 'true' transformants. The frequently-used SMGs such as antibiotics or herbicides generally obtained from prokaryotes, posing risks due to their lethal nature. In this regard, some safety SMGs based on visual, abiotic stress and metabolism are more attractive and successfully used for screening of transformants with high selection efficiency. This review mainly focus on their availability and applicable range which contributes to breeding biosafety transgenic plants.

Key words: transgenic plants; safety maker; biosafety

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